

# Monitoring biodiversity using environmental DNA: the future is already here.

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**ID-GENE**  
ecodiagnosics

Research Symposium SNP+ in Zernez, June 2, 2023



# Environmental DNA (eDNA)

All the **genetic material (DNA)** from living organisms or their traces (skin cells, mucus, scales, urine, feces, saliva, gametes or decomposing bodies) present in the environment (water, soil, biofilm, etc.)

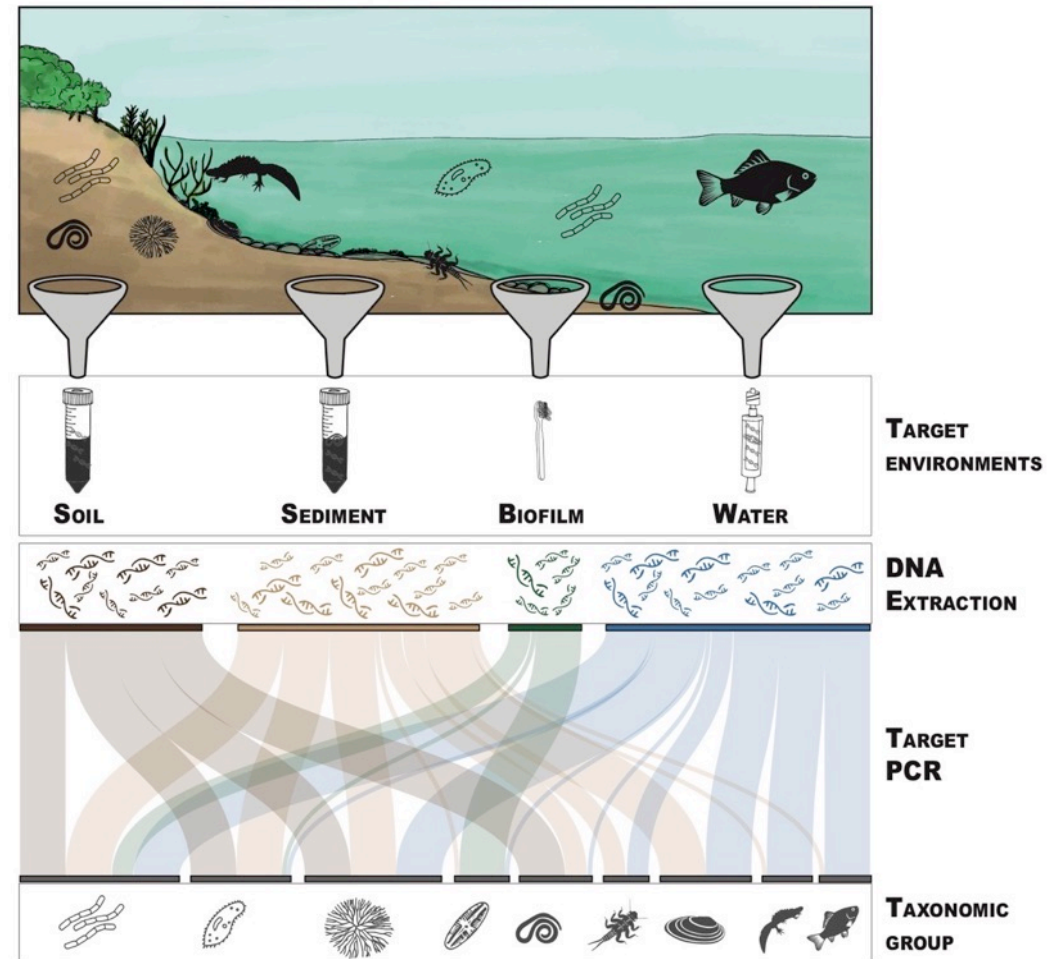
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NEWS AND VIEWS  
OPINION

MOLECULAR ECOLOGY WILEY

**Environmental DNA: What's behind the term? Clarifying the terminology and recommendations for its future use in biomonitoring**

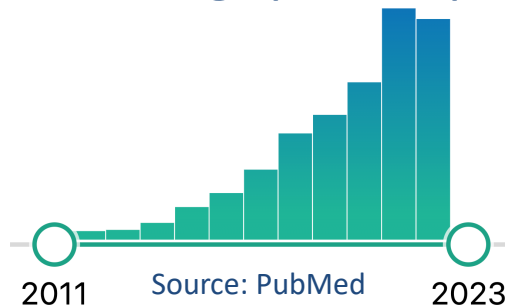
Jan Pawłowski<sup>1,2,3</sup> | Laure Apothéloz-Perret-Gentil<sup>1,2</sup> | Florian Altermatt<sup>4,5</sup>



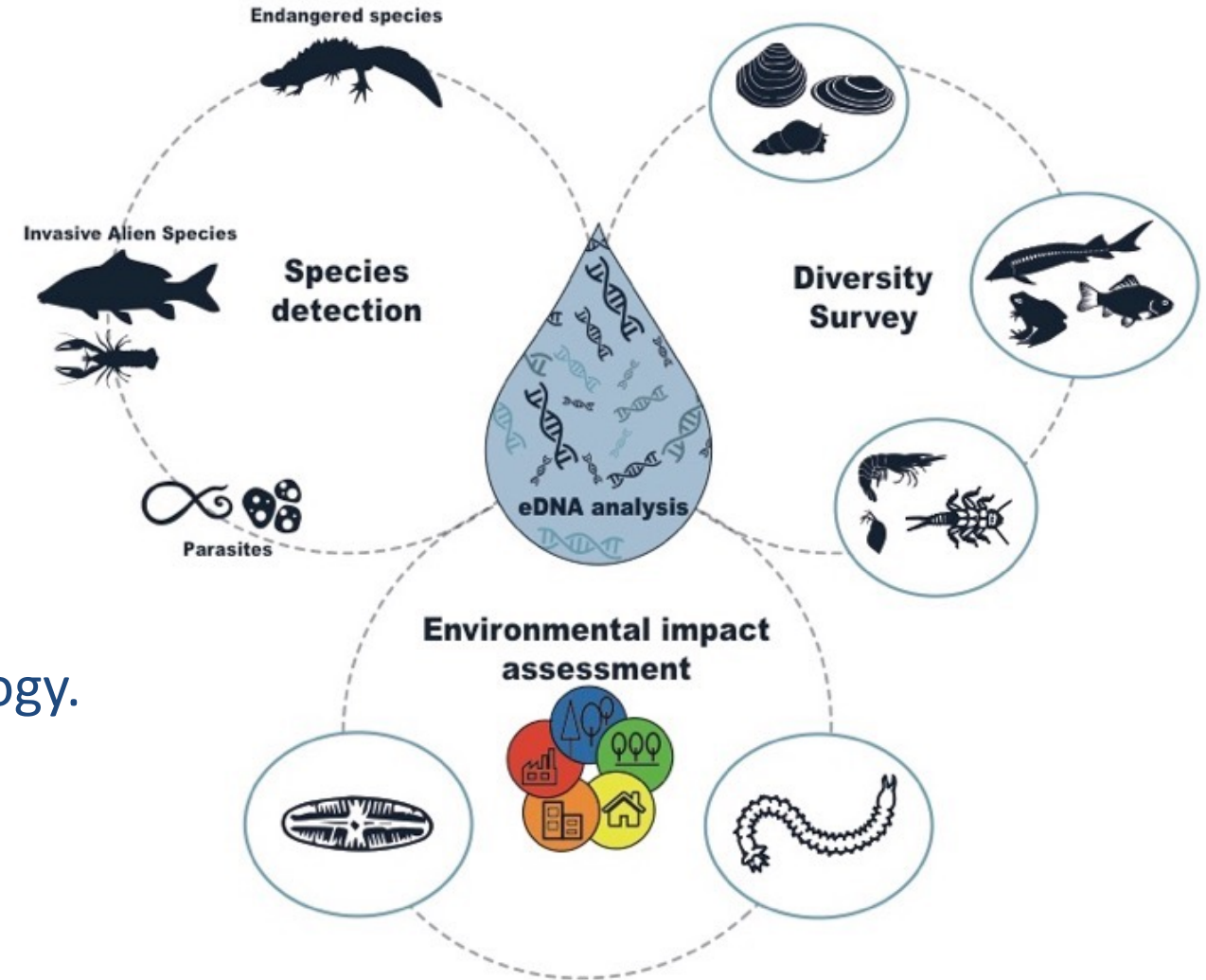
# eDNA applications

- Species detection and identification
- Biomonitoring and biodiversity survey
- Environmental impact assessment (biotic indices)

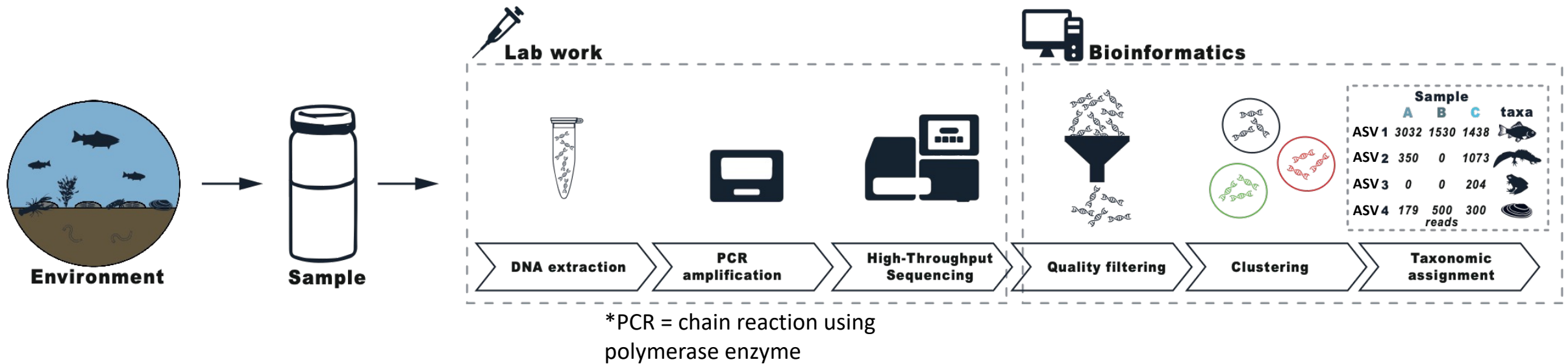
**Metabarcoding** approach uses high-throughput sequencing technology.



Number of publications on metabarcoding, per year



# Metabarcoding workflow



Pre- and post-amplification laboratory analyses are conducted in separated rooms.

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Extraction



PCR




Post-PCR



Sequencing

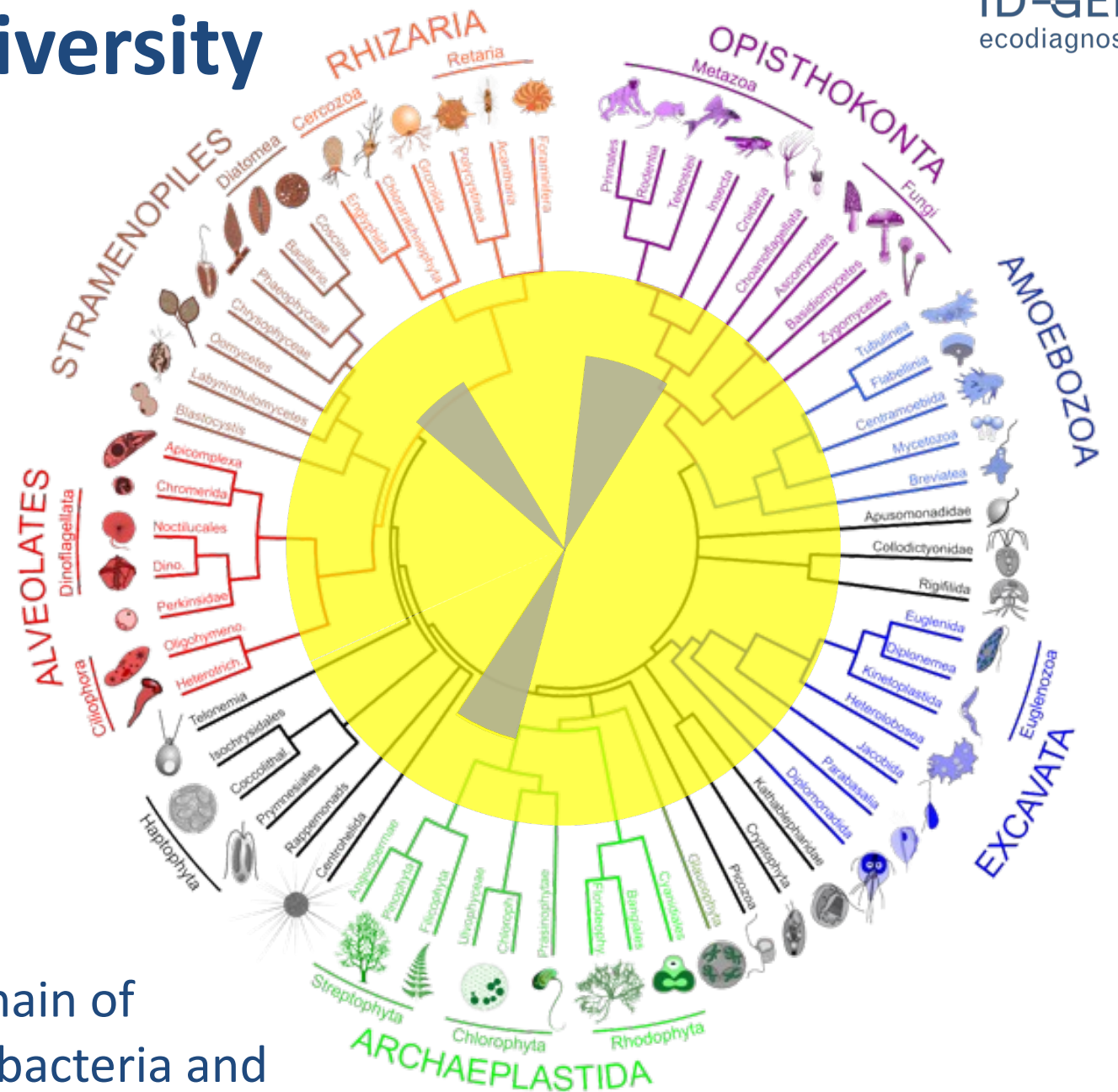
# Holistic approach to biodiversity

The same environmental sample allows to study eDNA of all the organisms of the Tree of life, from bacteria to animals

 groups studied with the Biology module of the Swiss system for analysing watercourses

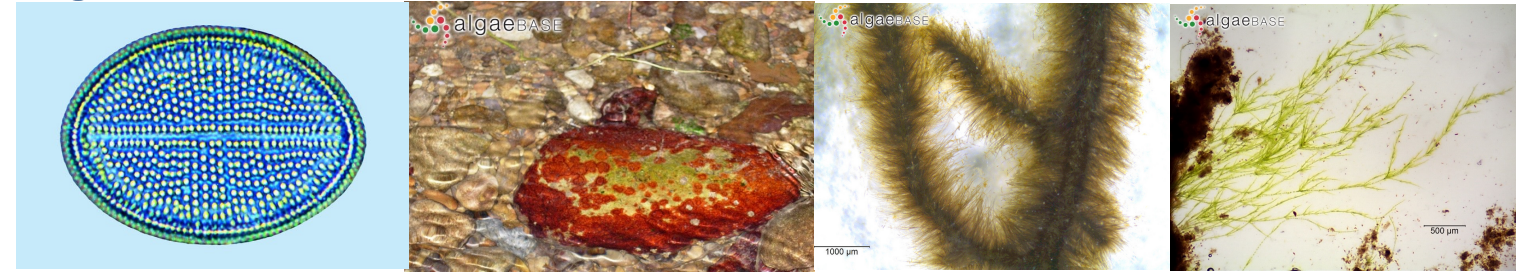
 groups studied with the eDNA approach

Tree of life of the domain of eukaryotes (domains bacteria and archea are not represented)

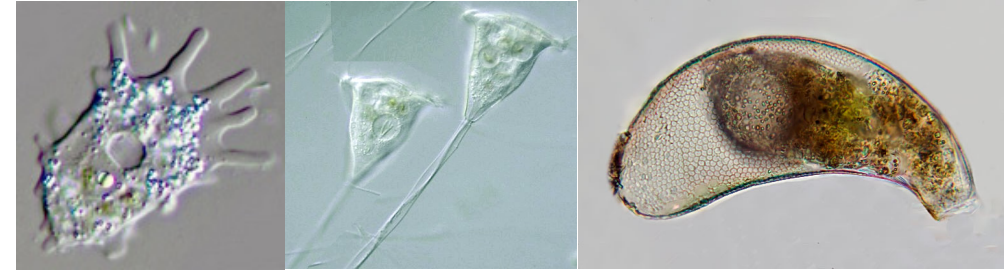


# Holistic biodiversity inventories

## Algae



## Protists



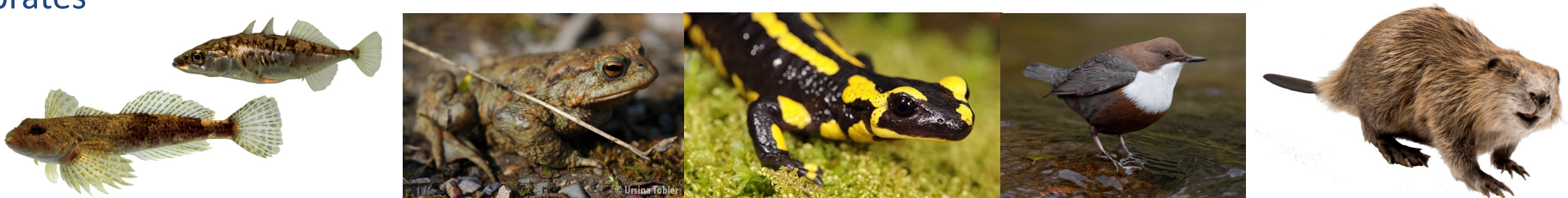
## Meio-/micro-invertebrates



## Macro-invertebrates



## Vertebrates



# Fish community survey

| Species                          | Common name              | Status CH             | Drize | Aire | Allondon | Vengeron | Versoix | Seymaz |
|----------------------------------|--------------------------|-----------------------|-------|------|----------|----------|---------|--------|
| <i>Salmo labrax</i>              | Black Sea salmon         | Critically Endangered | X     |      | X        |          | X       | X      |
| <i>Thymallus thymallus</i>       | European grayling        | Endangered            |       |      | X        |          | X       |        |
| <i>Alburnoides bipunctatus</i>   | Spirlin                  | Vulnerable            | X     |      |          |          |         | X      |
| <i>Telestes souffia</i>          | Souffia                  | Vulnerable            |       |      |          |          |         | X      |
| <i>Barbatula barbatula</i>       | Stone loach              | Near Threatened       |       | X    | X        |          | X       | X      |
| <i>Barbus barbus</i>             | Common barbel            | Near Threatened       |       |      | X        |          | X       | X      |
| <i>Cottus gobio</i>              | European bullhead        | Near Threatened       |       |      | X        |          | X       |        |
| <i>Salmo trutta/trutta fario</i> | Brown trout              | Near Threatened       | X     | X    | X        |          | X       | X      |
| <i>Esox lucius</i>               | Northern pike            | Least Concern         |       |      |          |          | X       |        |
| <i>Gobio gobio</i>               | Gudgeon                  | Least Concern         |       | X    | X        |          | X       | X      |
| <i>Perca fluviatilis</i>         | European perch           | Least Concern         | X     |      |          |          | X       | X      |
| <i>Phoxinus phoxinus</i>         | Common minnow            | Least Concern         | X     | X    | X        | X        | X       | X      |
| <i>Rutilus rutilus</i>           | Common roach             | Least Concern         | X     | X    | X        |          | X       | X      |
| <i>Squalius cephalus</i>         | European chub            | Least Concern         | X     | X    | X        |          | X       | X      |
| <i>Tinca tinca</i>               | Tench                    | Least Concern         |       |      |          |          |         | X      |
| <i>Ameiurus nebulosus/melas</i>  | Brown/Black bullhead     | Introduced            | X     |      |          |          | X       | X      |
| <i>Gasterosteus aculeatus</i>    | Three-spined stickleback | Introduced            |       |      |          |          | X       |        |
| <i>Lepomis gibbosus</i>          | Pumpkinseed              | Introduced            | X     |      |          |          | X       | X      |
| <i>Oncorhynchus mykiss</i>       | Rainbow trout            | Introduced            | X     | X    |          |          | X       |        |



Black Sea salmon



European Grayling



Spirlin



Souffia



Common barbel

## Congruence of eDNA and conventional inventories:

|          | eDNA (2022) | conventional method |
|----------|-------------|---------------------|
| Vengeron | 1 species   | 0 species in 2021   |
| Seymaz   | 13 species  | 13 species in 2019  |

# Protected species survey

Metabarcoding allows to detect all species of newts in Switzerland.



Lissotriton helveticus



Lissotriton vulgaris



Lissotriton vulgaris  
meridionalis



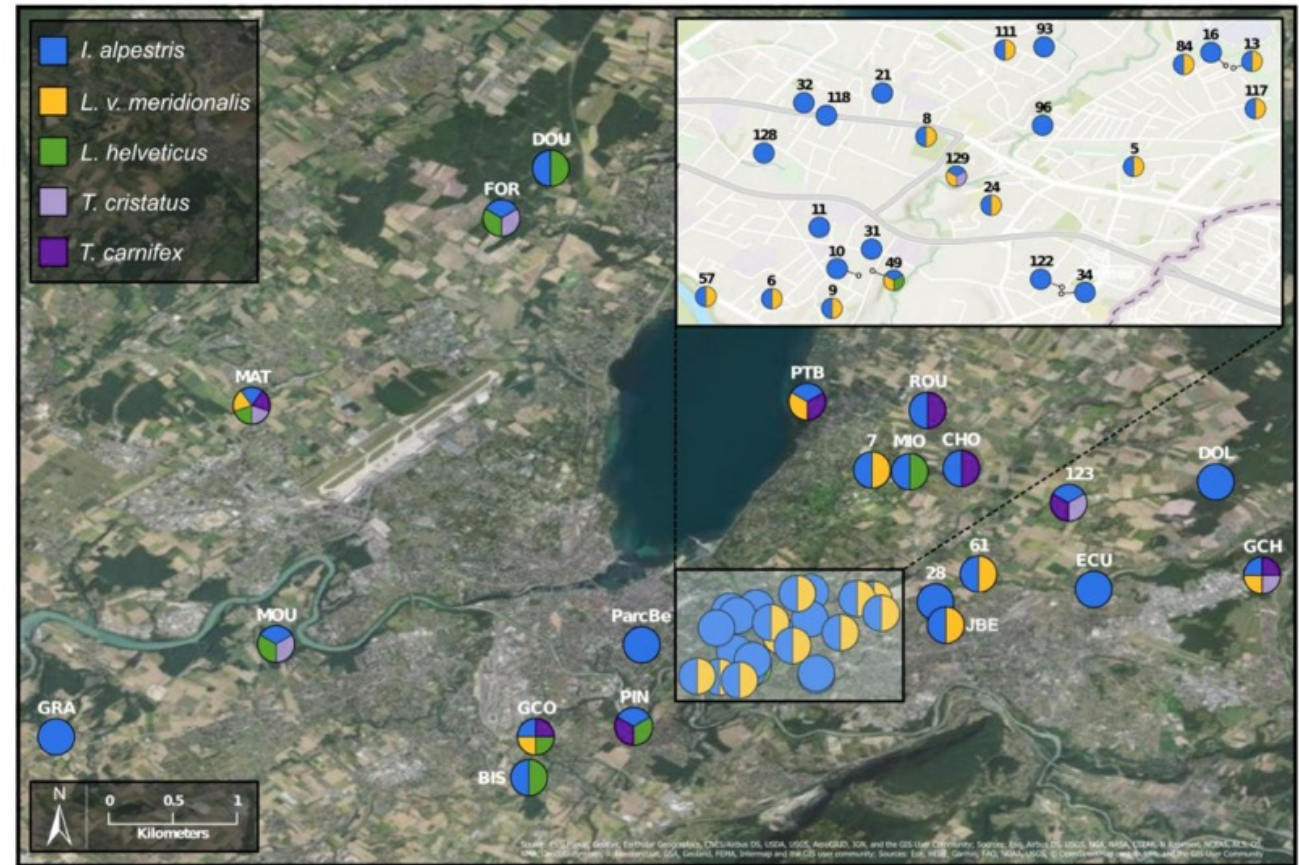
Ichthyosaura  
alpestris



Triturus  
cristatus/carnifex

## Monitoring newt communities in urban area using eDNA metabarcoding

Léo Charvoz<sup>1</sup>, Laure Apothéloz-Perret-Gentil<sup>1,2</sup>, Emanuela Reo<sup>1</sup>,  
Jacques Thiébaud<sup>3</sup> and Jan Pawlowski<sup>1,2,4</sup>

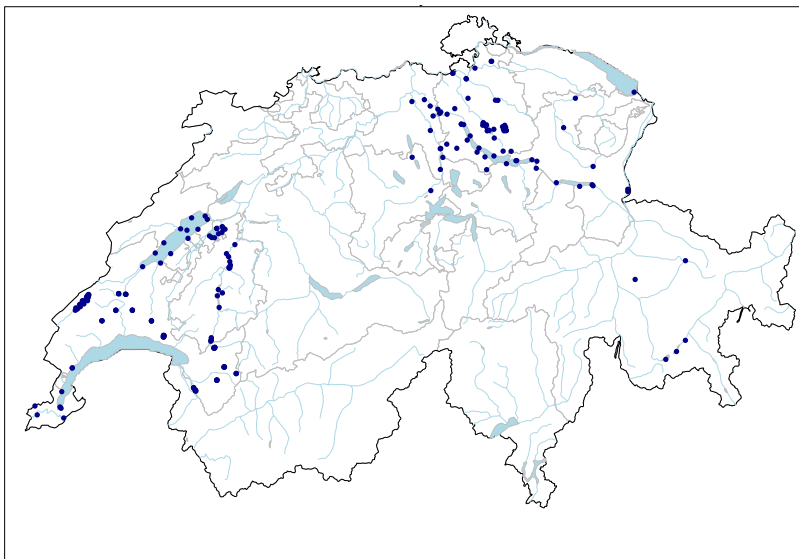


# Invasive species detection

**Invasive species monitoring**  
is recommended by Swiss Federal  
Office for the Environment and  
conducted by the cantons.

ID-Gene projects:

- Canton Vaud
- Canton Zürich
- Canton St. Gall
- Canton Argovie
- Canton Grisons
- Canton Geneva



*Dreissena bugensis*



*Corbicula fluminea*



*Neogobius melanostomus*



*Ponticola kessleri*



*Dikergammarus villosus*



*Crangonyx pseudogracilis*



*Physella acuta*



*Pacifastacus leniusculus*

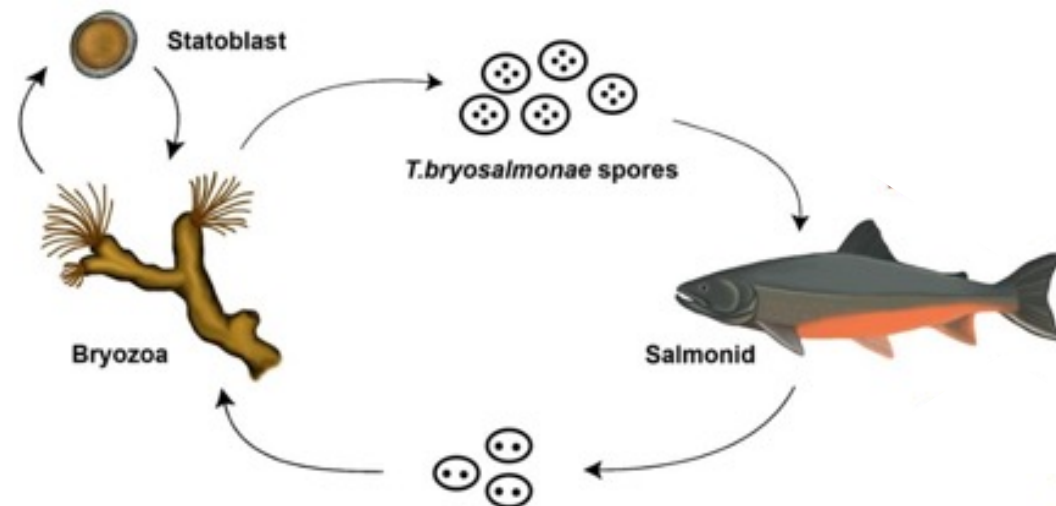
# Pathogen detection in water samples

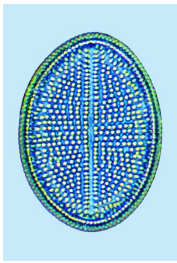
## Crayfish pathogens:

- Oomycete *Aphanomyces astaci* causing the Crayfish plague

## Fish pathogens:

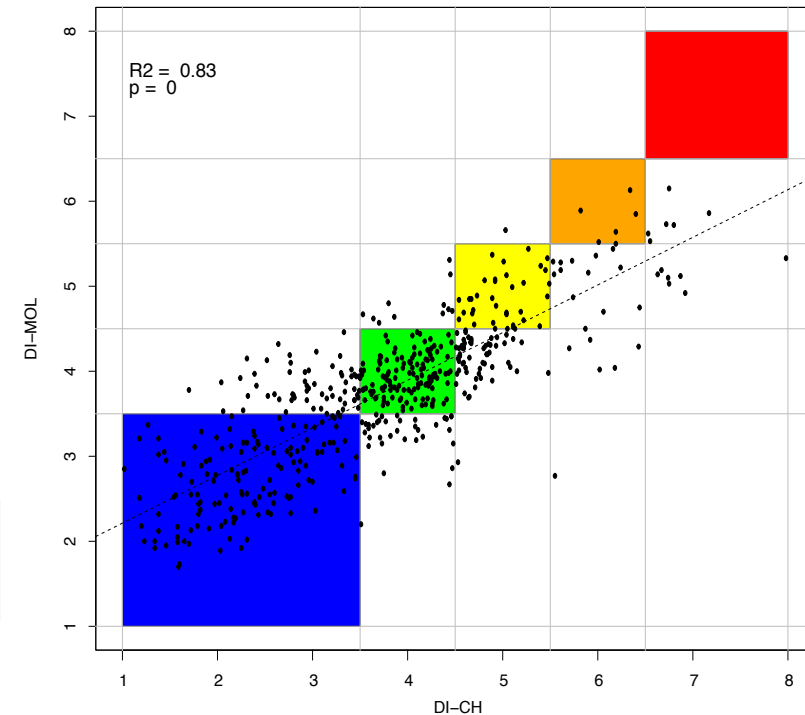
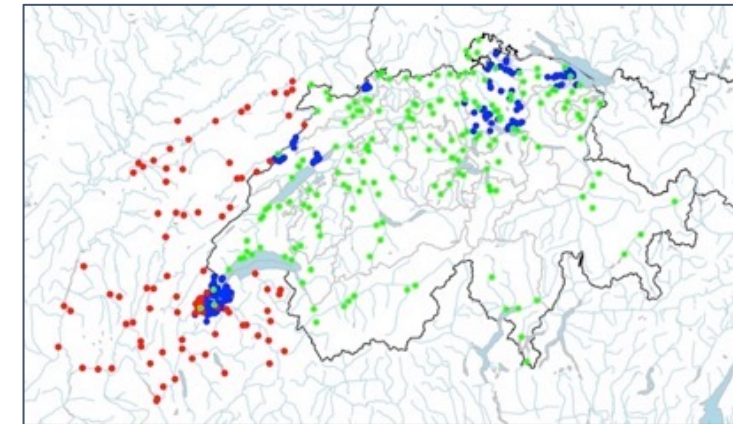
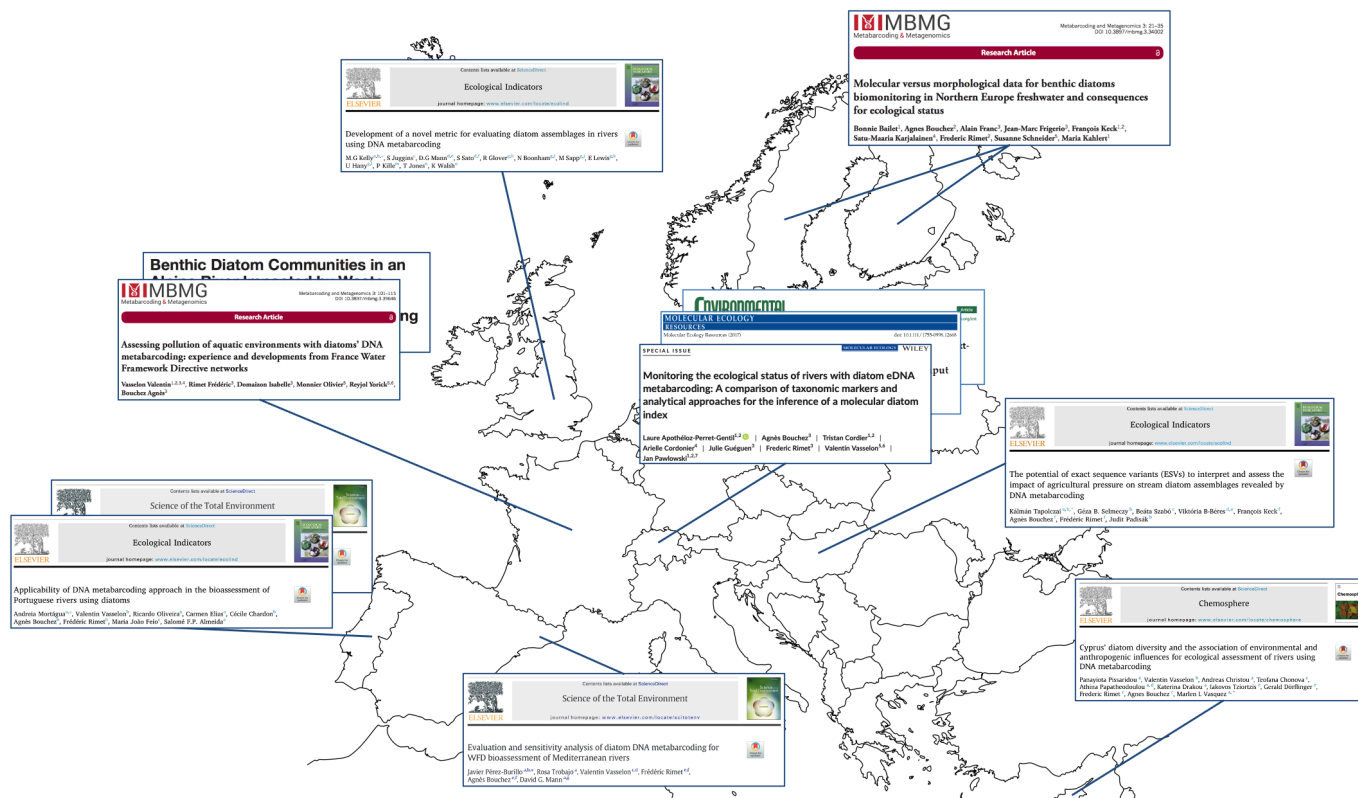
- Oomycete *Saprolegnia parasitica* causing saprolegniasis
- Cnidarian *Tetracapsuloides bryosalmonae* causing the Proliferative kidney disease (PKD)





# Diatom Molecular Index

DNA-based diatom molecular index has been successfully developed in Switzerland and other European countries showing excellent correlation between molecular and morphological data.



**Authors:**

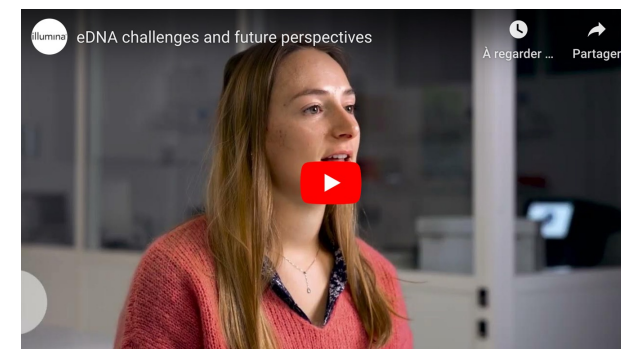
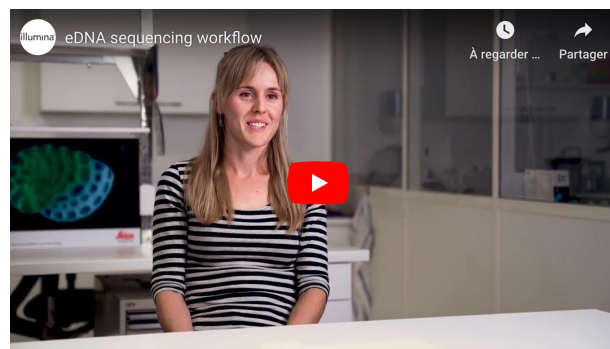
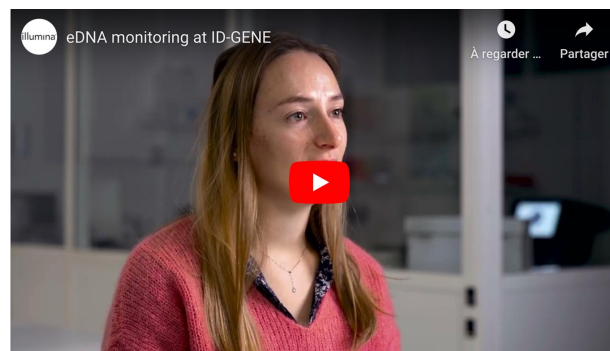
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**Advisory group:**

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illumina <https://ilmnmkt.illumina.com/2016011140>

- What is environmental DNA, or eDNA?
- How is eDNA analysis applied for biomonitoring of aquatic ecosystems?
- What are the steps of the eDNA sequencing workflow, from field to lab?
- What are the current challenges and future directions for a wider implementation of eDNA sequencing in routine environmental monitoring?



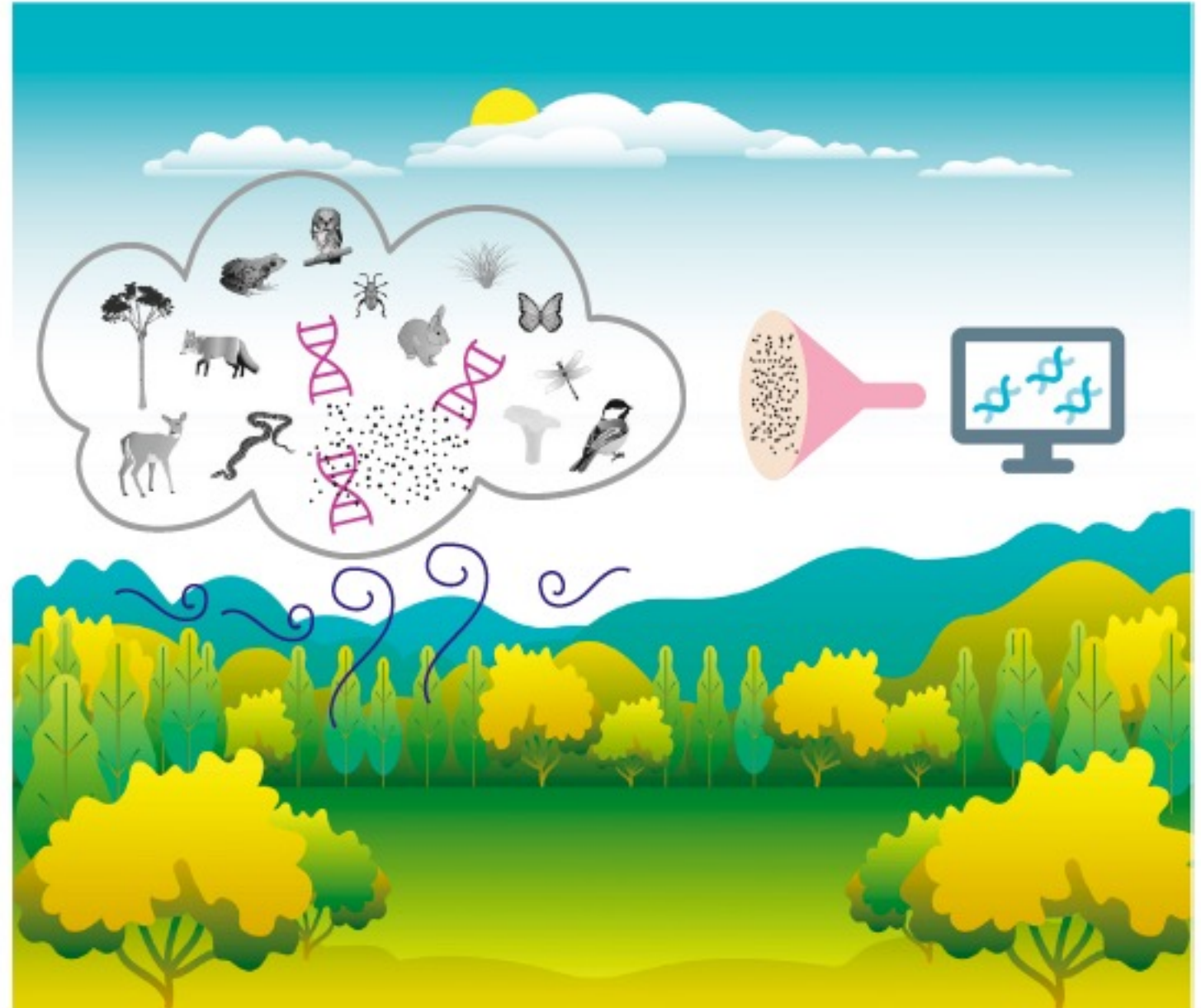
# Perspectives

## - Airborne eDNA

measuring biodiversity from DNA in the air

Pilot studies:

- Vertebrates (Lynggaard et al. 2022)
- Insects (Pumkaeo et al. 2021, Roger et al. 2022)
- Plants (Johnson et al. 2021)
- Microbes (de Groot et al. 2021, Gusareva et al. 2022)

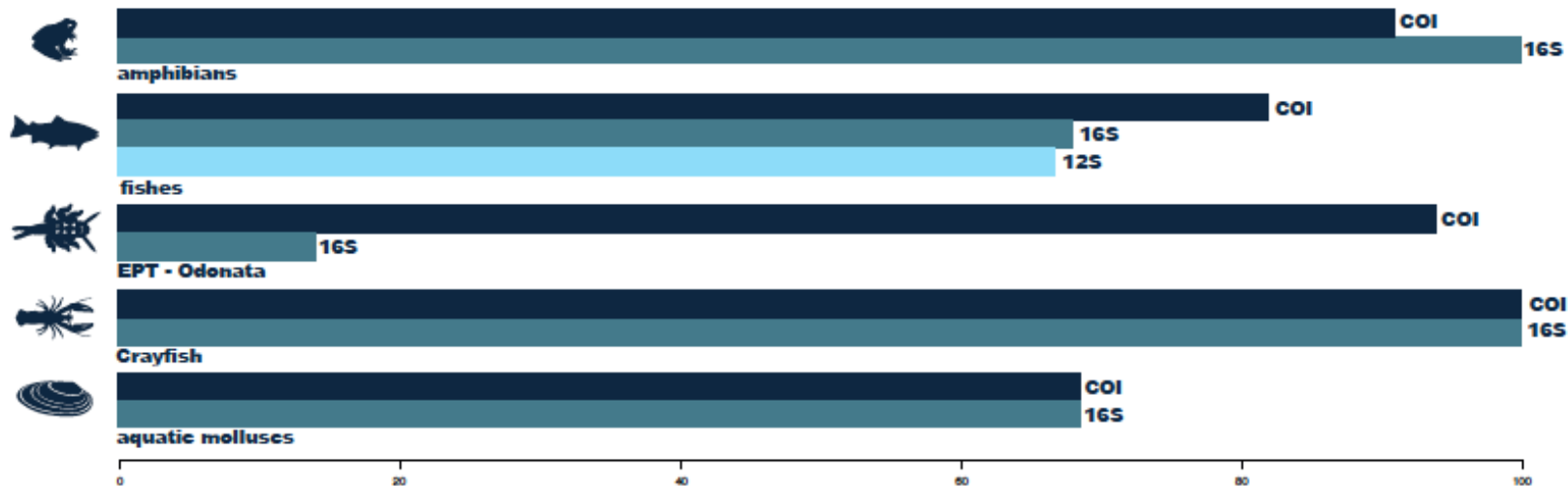


# Challenges and limitations

Completing barcoding reference databases



**Swiss Barcode of Life**  
A genetic inventory of Swiss biodiversity



eDNA does not allow:

Inferring species abundance

Detecting changes of any morphological features

Evaluating physiological state (population age)

Detecting hybridization

# Advantages of eDNA



Easy and non-invasive sample collecting without toxic fixative substances



Processing in batches using standardized protocols and automated processes – volume, reproducibility



Fast pipeline - results within few weeks



Sensitivity – early detection and detection of rare or elusive species



Biotic indices can be inferred from molecular data, congruency with morphological approach

## Conclusion

Metabarcoding approach allows processing of higher numbers of samples and thus allows better coverage and higher frequency of monitorings compared to morphological analysis.

Whole biodiversity from bacteria to animals can be assessed simultaneously from a sample.

➤ Metabarcoding is a robust ready-to-use tool with potential to improve biomonitoring.

# Our partners in Switzerland

Schweizerische Eidgenossenschaft  
Confédération suisse  
Confederazione Svizzera  
Confederaziun svizra



Bundesamt für Umwelt BAFU  
Office Fédéral de l'environnement OFEV  
Ufficio federale dell'ambiente UFAM  
Uffizi federal d'ambient UFAM

**eawag**  
aquatic research ooo



SWISS NATIONAL SCIENCE FOUNDATION

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ÉCOLE POLYTECHNIQUE  
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DE GENÈVE



Kanton St.Gallen



Kanton Graubünden  
Chantun Grischun  
Cantone dei Grigioni



ETAT DE FRIBOURG  
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POST TENEBRAS LUX

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ET CANTON  
DE GENEVE



Kanton Zürich



KANTON AARGAU



Kanton Bern  
Canton de Berne



Kanton Basel



Kanton Zug



AquaPlus

La Maison  
de la Rivière

The logo for ID-GENE ecodiagnosics is positioned in the top right corner. It features the text "ID-GENE" in a large, bold, blue sans-serif font, with a horizontal line through the middle of the "G". Below it, the word "ecodiagnosics" is written in a smaller, lowercase, blue sans-serif font. The background of the logo is a light blue sky with soft white clouds.

**ID-GENE**  
ecodiagnosics

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**THANK YOU FOR  
YOUR ATTENTION!**

The bottom right section of the slide features a vibrant underwater scene. A bright rainbow arches across the surface of the water, which is filled with bubbles and splashes. Below the surface, various microscopic organisms, including bacteria and viruses, are depicted in a teal-colored environment. Overlaid on this scene is the text "INNOVATIVE DNA SOLUTIONS FOR ENVIRONMENTAL MONITORING" in a bold, white, sans-serif font.

**INNOVATIVE  
DNA SOLUTIONS  
FOR ENVIRONMENTAL  
MONITORING**